Effects of He-O₂ breathing during experimental decompression sickness following air dives

P. W. CATRON, L. B. THOMAS, E. T. FLYNN, JR., J. J. McDERMOTT, and M. A. HOLT

Hyperbaric Medicine Program Center, Naval Medical Research Institute, Bethesda, MD 20814-5055

Heliun - diator

Catron PW. Thomas LB. Flynn ET Jr. McDermott JJ. Holt MA. Effects of He-O₂ breathing during experimental decompression sickness following air dives. Undersea Biomed Res 1987; 14(2):101-111.—The effects of ventilation with He-O₂ during decompression sickness (DCS) and venous air embolism were studied. Fifteen anesthetized dogs were mechanically ventilated and subjected to repeated air dives until pulmonary artery pressure at least doubled within 10 min postdive. At 30 min postdive, ventilation was either continued with air (controls, n = 7) or changed to He-O₂ (n = 8) for an additional 90 min. All animals developed pulmonary hypertension, systemic hypotension, hemoconcentration, hypoxemia, hypercarbia, and pulmonary edema. Breathing air or He-O₂ postdive did not alter these responses, but He-O₂ breathing produced an 11% increase in pulmonary vascular resistance (PVR). In 3 other anesthetized dogs that were not subjected to dives, ventilation was changed to He-O₂ at various times during an intravenous infusion of air: He-O₂ breathing caused a 22% increase in PVR. We conclude that breathing He-O₂ during DCS resulting from an air dive can intensify pulmonary vascular obstruction.

diving decompression sickness venous air embolism helium-oxygen chokes pulmonary edema

animals

Decompression sickness (DCS) is a well-established hazard of air diving. The use of He-O₂ mixtures for treatment of DCS resulting from an air dive has been previously advocated, but the proposed benefits are supported only by anecdotal evidence (1-5). Due to the different diffusivities of helium and nitrogen, breathing He-O₂ in air DCS could theoretically pose a significant hazard. Transient bubble growth resulting from a more rapid diffusion of helium into the bubble than nitrogen out of it could materially worsen symptoms.

The central location of the lung within the circulation makes it a major target organ in DCS. Bubble embolization of the lung during DCS regularly results in pulmonary hypertension and impaired gas exchange (6-8). These responses might be expected to increase if breathing He-O₂ caused growth of nitrogen-filled bubbles within the

DISTRIBUTION STATEMENT A

Approved for public release;

pulmonary circulation. This hypothesis is supported by the experimental observation that ventilation of animals with $He-O_2$ during venous air embolism worsens pulmonary hypertension and hypoxemia (9). To test this hypothesis, we studied the effects of $He-O_2$ breathing in an animal model of DCS.

MATERIALS AND METHODS

Nineteen experiments were completed using male mongrel dogs. The dogs were divided into 3 groups, an air control group (n = 7), an He-O₂ group (n = 8), and an air embolism group (n = 3). Mean weights $(\pm \text{ SD})$ of each group were: air controls $12.6 \pm 0.5 \text{ kg}$; He-O₂ $16.2 \pm 2.6 \text{ kg}$; and air embolism group $12.3 \pm 0.8 \text{ kg}$.

Fasted animals were anesthetized with a loading dose of sodium pentobarbital (30 mg/kg, i.v.), endotracheally intubated, and placed on a mechanical ventilator breathing air. Anesthesia was maintained by a continuous intravenous infusion of sodium pentobarbital (0.068 mg · kg⁻¹ · min⁻¹), supplemented with intravenous boluses of 50 mg as needed to suppress corneal reflex and spontaneous ventilation.

A pressure-cycled ventilator (Bird Products, Palm Springs, CA) was used during surgical preparation. At all other times a constant-volume ventilator (Penlon Ltd, Abingdon, England) was used. Before the predive data collection the ventilator was adjusted to provide physiologic values of arterial pH, oxygen partial pressure (PCO₂), and carbon dioxide partial pressure (PCO₂). After this adjustment no further change in ventilator volume or rate was made during the experiment. The oxygen content of the He-O₂ mixture used in the He-O₂ group was measured using a Beckman OM-11 oxygen analyzer (Beckman Corp., Schiller Park, IL). The inspired He-O₂ mixtures always contained between 20.5 and 20.9% oxygen.

A 7-French Swan-Ganz catheter was advanced through a femoral vein into the pulmonary artery to measure pulmonary artery pressure and cardiac output (CO). For the measurement of systemic arterial pressure, a 1.77-mm i.d., fluid-filled catheter was advanced through one femoral artery into the midthoracic aorta. To measure left ventricular end-diastolic pressure (LVEDP), a 0.96-mm i.d., fluid-filled catheter was placed in the other femoral artery and advanced retrograde across the aortic valve into the left ventricle. Vascular pressures were measured using Gould P23 pressure transducers and Gould pressure processors (Gould, Cleveland, OH). All vascular pressures were referenced to midchest level. CO was measured using a thermodilution cardiac-output computer (Edwards Laboratories, Santa Ana, CA). Mean pulmonary artery pressure (PAP), CO, and LVEDP were used to calculate pulmonary vascular resistance (PVR). All animals were instrumented with electrocardiographic leads, a rectal temperature probe, and a urinary bladder catheter.

Baseline predive measurements were made of core body temperature, CO, hematocrit (Hct), arterial PO₂, PCO₂, and pH, and vascular pressures. In animals of the He-O₂ group, predive measurements were made of all variables during ventilation with air and with He-O₂. After baseline measurements, the vascular catheters were filled with a volume of heparinized saline (5 U/ml) equivalent to catheter volume, and animals were subjected to a simulated air dive to 10 ATA in an animal recompression chamber (Hahn and Clay, Houston, TX). For all dives the descent rate to 10 ATA was 3.0 ATA/min (3-min descent). The ascent rate from 10 ATA was 1.8 ATA/min (5-min ascent). For the first dive the time at 10 ATA was always 14 min. After the

animals had returned to the normal surface pressure of 1 ATA the systolic PAP was monitored for 10 min. If systolic PAP at least doubled during this time, no further dives were made. If lesser pressure increases occurred, repetitive dives each with a 3-min descent, 5 min at 10 ATA, and a 5-min ascent were made until systolic PAP at least doubled during the postdive observation period of 10 min.

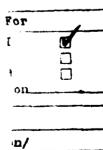
Postdive measurements of physiologic variables were made at 15, 45, 75, and at 105 min after surfacing. These measurements included core temperature, Hct, arterial PO2, PCO2, and pH, CO, and vascular pressures. Thirty minutes after surfacing, the inspired gas was either changed to He-O₂ (He-O₂ group) or mechanical ventilation was continued using air (air control group). Vascular pressures were recorded continuously for 10 min before and after switching to the test gas. CO was measured every 1-2 min for 5 min before and for 10 min after gas switching.

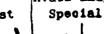
At 120 min postdive, the anesthetized animals were killed with an intravenous bolus of KCI. The lungs were removed, inflated with 10% buffered formalin solution, and prepared for routine histologic examination.

In 3 additional animals (air embolism group), gas switching was studied during venous air embolism at 1 ATA. These animals were anesthetized and instrumented as in the decompression experiments, except that an additional catheter (1.66 mm i.d.) was inserted into the inferior vena cava for infusion of air. In 2 animals, air was infused intravenously for 40 min at a dosage of 0.1 ml · kg⁻¹ · min⁻¹, a level that resulted in a stable mean PAP approximately two times the baseline value. At 30 min into the infusion, ventilation was changed to He-O₂ for 10 min while air infusion was continued. In the 3rd animal, air was infused at a rate of 0.35 ml · kg⁻¹ · min⁻¹ to raise mean PAP to approximately three times baseline values. In this animal ventilation with He-O₂ was begun after 13 min of air embolism and continued for 6.5 min, when it was switched back to air and the air infusion ended. At the end of the air embolism experiments, animals were killed and their lungs removed for histologic examination in the manner described for the decompression experiments.

Statistical analyses were done using either a paired t test or an analysis of variance. A paired t test was used to compare baseline values of vascular pressures and arterial blood gases during ventilation with air and He-O2 and to compare mean PAP and PVR before and after gas switching. Two analysis of variance procedures were used. Differences in PVR before and after gas switching were examined using a model that estimated a preswitch value of PVR for each animal and a single percentage increase for the entire group of animals after gas switching. Residual error from an eightparameter model (no increase postswitch) was compared to that from a nine-parameter model (an equal percentage increase for all animals). Another analysis of variance procedure was used to test the hypothesis that with regard to a given physiologic measurement, control and dived animals were different at one or more times. Physiologic data were fit to a five-parameter model that estimated one mean value for the predive baseline and a mean value at each of the postdive measurement times. The residual error from this model was compared with that from a 10-parameter model in which pre- and postdive means were estimated for both control and dived groups. In both analyses of variance, means were estimated by a least squares method using a FORTRAN version of the iterative parameter estimation program of Bailey and Homer (10). Residual errors were compared using an F test, and an alpha level of 1ty Codes 0.05 was taken as an indication of statistical significance. Avail and/or







RESULTS

Responses to decompression

The dive profile used in these experiments caused responses that are associated with stressful decompression: pulmonary hypertension, systemic hypotension, decreased CO, systemic acidosis, hypoxemia, hypercarbia, hemoconcentration, and decline in body temperature (6-8). Figures 1 and 2 show that although PAP, mean arterial pressure (MAP), CO, Hct, pH, PO₂, PCO₂, and body temperature all reflected the deleterious effects of the dive, there were no important effects of a switch to He-O₂ (compare broken and solid lines to the right of the vertical).

Body temperatures (Fig. 2d) of animals of the air group were lower than those of the He-O_2 group, and the differences between the 2 groups were statistically significant (P < 0.001). The differences were present predive and remained relatively constant throughout the experiment. The lower temperatures of animals in the air group presumably reflected a more rapid loss of body heat under general anesthesia. This relative inability to maintain thermoregulation may have been the result of a higher ratio of body surface area to mass, because animals in this group weighed less (mean weight 12.6 kg) than those of the He-O₂ group (mean weight 16.2 kg).

Hemodynamic effects

In animals of the He-O₂ group, predive values of mean PAP, PVR, MAP, CO, arterial pH, PO₂, and PCO₂ were not significantly different during ventilation with He-O₂ or air. The hemodynamic consequences of changing ventilation from air to He-O₂ during DCS and air embolism are summarized in Fig. 3a-d. These figures depict for each animal the average values of mean PAP or PVR for 3 min before and 4 min after the switch to He-O₂. (PVR is expressed in peripheral resistance units [PRU].) One PRU is equal to a pressure drop of 1 mmHg occurring with a flow of 1 ml·s⁻¹. (Normal PVR is approximately 0.2 PRU.) In the decompression experiments, no significant change in mean PAP was seen after the switch to He-O₂ (Fig. 3a). PVR, however, rose in these animals from an average of 1.18 PRU to 1.30 PRU after switching to He-O₂, an increase of 11% (Fig. 3b). This increase was statistically significant by both paired t test and analysis of variance (P < 0.05). The increase in PVR with switching was small when compared with the increase produced by the dive itself. After the dive, average PVR rose from 0.17 to 1.176 PRU, an increase of approximately 700%.

By contrast to the decompression studies, during venous air embolism, changing the gas mixture from air to He-O₂ increased mean PAP by an average of 7 mmHg within 4 min of the switch (Fig. 3c). PVR rose in these animals by an average of 0.20 PRU after switching to He-O₂, an increase of 24% (Fig. 3d).

Increases in PVR and mean PAP after the shift to He-O₂ were much easier to demonstrate in animals with venous air embolism than in those with DCS. Although the sample size was too small for statistical correlations, these differences did not seem to be related to the preexisting level of PVR or CO. In animals with DCS the average preswitch CO was 0.157 liter · min⁻¹ · kg⁻¹ and the average preswitch PVR was 1.18 PRU. Changing to He-O₂ increased mean PAP by 1 mmHg and PVR by 0.12 PRU (11%). The 1 animal given intravenous air at a rate sufficient to increase baseline mean PAP threefold had preswitch levels of CO of 0.170 liter · min⁻¹ · kg⁻¹ and PVR

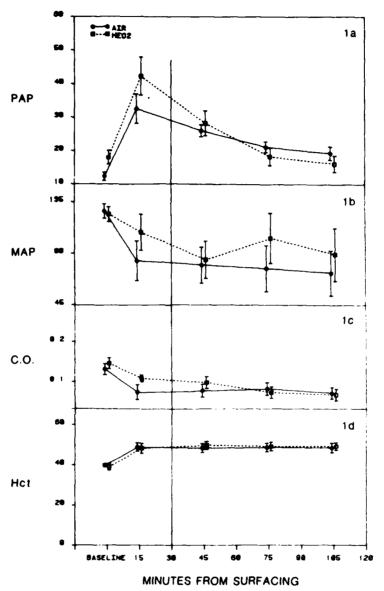


Fig. 1. Mean values of hemodynamic variables during DCS in animals of the air and He-O₂ groups. a, Mean pulmonary artery pressure mmHg; b, mean systemic arterial pressure mmHg; c, cardiac output, liter \cdot min $\frac{1}{2} \cdot \text{kg}^{-1}$; d, hematocrit. In all figures error bars denote \pm 1 sE and vertical line indicates time of switch to He-O₂ breathing.

of 1.26 PRU. Changing to ventilation with He- O_2 in this animal increased mean PAP by 8 mmHg and PVR by 0.29 PRU (22%). Thus, although this animal was comparable to those with DCS in terms of preswitch CO and PVR, switching to He- O_2 caused a substantially greater degree of pulmonary vascular obstruction.

Pathologic changes within the lungs were similar for animals in the air and He-O₂ groups. No animal in either group had normal lungs. The most prominent abnormality was the presence of pulmonary edema. Almost all animals of both groups had cuffs

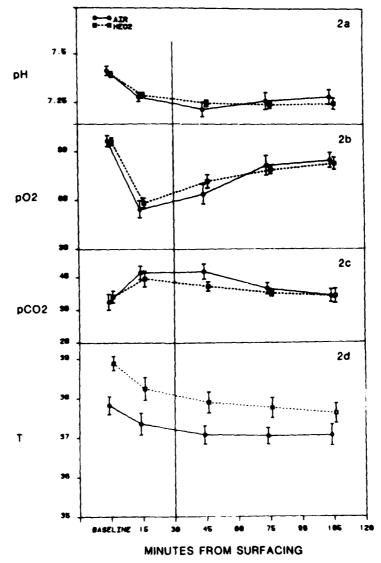


Fig. 2. Mean values of arterial blood gas variables and temperature during DCS in the air and He-O₂ groups. a, Arterial pH; b, arterial PO₂ mmHg; c, arterial PCO₂ mmHg; d, temperature °C. Error bars denote \pm 1 SE and vertical line indicates time of switch to He-O₂ breathing.

of edema fluid encircling pulmonary vessels and bronchi, and some animals in each group had evidence of alveolar filling. There was no apparent difference in severity of the pathologic findings between the two groups.

One animal in the air embolism group had normal lungs. The lungs of the other 2 showed perivascular and peribronchial cuffs of edema fluid. Alveolar filling was not seen in the lungs of any of these animals.

DISCUSSION

Helium-oxygen has been proposed as a suitable breathing mixture for use during recompression therapy of DCS that has resulted from an air dive (1-5). Usually, no

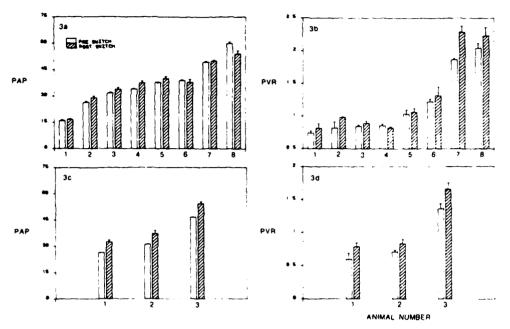


Fig. 3. Mean PAP mmHg (a) and PVR (PRU) (b) for each animal before and after switching to He-O₂ during DCS. Mean PAP mmHg (c) and PVR (PRU) (d) for each animal before and after switching to He-O₂ during venous air embolism. Error bars denote ± 1 SE.

specific rationale has been offered, and the recommendation implies that $He-O_2$ is an acceptable substitute for air if its use is operationally convenient. It has also been suggested that $He-O_2$ might be beneficial in cases of DCS with labored breathing (11) or that it might provide a favorable gradient for the elimination of nitrogen (3). James (12) recommended that $He-O_2$ be used as a treatment gas for all recompression treatments in lieu of both oxygen and air. He cites lowered breathing resistance and rapid elimination of nitrogen as advantages of $He-O_2$ gas mixtures.

If a breathing gas has a higher solubility or diffusivity in blood than nitrogen, the risk exists that intravascular bubbles containing nitrogen may temporarily grow through a counterdiffusion mechanism. Such bubble growth might be expected to worsen vascular obstruction.

In the case of bubbles in the pulmonary circulation, the lung may be viewed as an infinite source for inward diffusion of a new gas into intravascular bubbles and an infinite sink for outward diffusion of gas from within bubbles. The effect of gas solubility and diffusivity on changes in bubble size can be appreciated by examination of Fick's law of diffusion expressed in spherical coordinates:

$$dV/dt = \alpha DA dP/dr$$

where V is the volume of inert gas, α the solubility and D the diffusivity of the inert gas in blood, P the partial pressure of the inert gas in blood, and r the bubble radius (13).

Assuming that the parameters A and r are the same for the two gases, and P is equal but of opposite sign, the net effect on bubble size of changing breathing gases during DCS will depend on the relative products of solubility and diffusivity of the two gases $(\alpha_1 D_1/\alpha_2 D_2)$.

The effect of relative solubility is well illustrated in experiments where nitrous oxide is breathed during the course of venous air embolism. Nitrous oxide is approximately equal to nitrogen in diffusivity (in water at 25°C, $DN_2O = 2.10 \times 10^{-5}$ cm²/s, $DN_2 = 1.99 \times 10^{-5}$ cm²/s) (14) but is much more soluble (in blood at 37°C, $\alpha N_2O = 0.455$ ml N_2O/ml blood, $\alpha N_2 = 0.0158$ ml N_2/ml blood) (15). The ratio $(\alpha N_2ODN_2O/\alpha N_2DN_2)$ is approximately 30, and this should favor the growth of nitrogen-filled bubbles if the gas surrounding the bubble is abruptly changed to N_2O .

Some experimental observations support this analysis. Changing ventilation of animals from air to a mixture of nitrous oxide and oxygen during venous air embolism has been shown to increase pulmonary artery pressure and arterial PCO_2 , and to lower arterial PCO_2 , and to lo

A similar situation exists for CO_2 , which is much more soluble than nitrogen in blood. The ratio $\alpha CO_2 \cdot DCO_2/\alpha N_2 \cdot DN_2$ is approximately 38. Nitrogen-filled bubbles would be expected to grow when surrounded by CO_2 . Harvey (22) observed the behavior of nitrogen-filled bubbles as they rose through a water column successively layered with N_2 then CO_2 then N_2 saturated water. Bubbles rapidly enlarged when they encountered the CO_2 layer and shrank when they reencountered a nitrogen saturated layer.

Although He is less soluble than N_2 in blood (at 37° C, α He = 0.0104 ml He/ml blood, $\alpha N_2 = 0.0158$ ml N_2 /ml blood) (15), it is more diffusible in aqueous media than nitrogen (in water at 25° C, DHe = 6.2×10^{-5} cm²/s, DN₂ = 1.99×10^{-5} cm²/s). The ratio α HeDHe/ αN_2 DN₂ is approximately 2. Therefore, nitrogen-filled bubbles would be expected to grow if the gas surrounding the bubbles were abruptly changed from nitrogen to helium. Such an effect has been demonstrated under isobaric conditions with nitrogen-filled bubbles in gelatin (23) and seems to also occur in animals. Sergysels et al. (9) reported that ventilation of animals with He-O₂ during venous air embolism increased mean PAP and lowered arterial PO₂. These changes were thought to be caused by growth of intravascular bubbles within the pulmonary circulation due to rapid inward diffusion of helium.

We found that while He-O₂ breathing did produce a measurable increase in PVR, it did not cause a dramatic worsening of cardiopulmonary function. The most likely explanation for this lies in the severity of the model of DCS used in these experiments. This model corresponds most closely to a "blow-up" of a diver, a situation in which the unplanned ascent of the diver results in a large burden of omitted decompression and severe, usually fatal, DCS. Comparison of our data with that of Powell et al. (24) suggests that the bubble loads in our experiments were several times greater than those required to produce grade IV Doppler bubble signals. When compared to a 700% increase in PVR due to the dive profile used in these experiments, it is unlikely that an additional 11% increase resulting from gas switching would greatly worsen the existing severe impairment of pulmonary function. Although the relative degree of increase in PVR produced by changing ventilation to He-O₂ was small compared

to the dive, it was not small in absolute terms. The average measured increase in PVR produced by switching to He-O₂ was 0.12 PRU, a value which is 70% of the average predive baseline PVR of 0.17 PRU.

Venous gas embolism is not a complete model of DCS. Although the lung's response in the two conditions is similar in many ways, changing to He-O₂ breathing during venous air embolism produces a significantly greater increase in mean PAP and PVR than it does during DCS. This difference in response is not due to differences in severity of the measured pulmonary vascular obstruction before the beginning of He-O₂ ventilation. It may be due to differences in the size of the obstructed vessels in the two conditions. Although we did not measure the size of the obstructing bubbles in these experiments, it is likely that the obstructed vessels were smaller in the decompression experiments. The starting diameter of the air bubbles as they were infused in the air embolism experiments was probably near that of the internal diameter of the infusion catheter (1.66 mm). In a recent study of venous air embolism in which I-mm diameter air bubbles were infused, bubbles were found in the lung microvasculature with diameters ranging from 100-1000 µm (25). By comparison, many more small bubbles were probably present after decompression. In a previous study in which a resonant bubble detector was used with this DCS model, large numbers of 4-µm-sized bubbles were detected postdive (26). Although it is not possible to predict changes in the size of bubbles as they migrate through the venous system to the lungs, it is likely that the average size of the bubbles obstructing the lung was smaller in the DCS experiments than in the air embolism experiments. If this is true, it may be that many more vessels were completely obstructed during DCS than during air embolism. Growth of bubbles would not be expected to materially worsen vascular obstruction when the bubbles were already totally obstructing small pulmonary vessels. On the other hand, if much of the preswitch pulmonary vascular resistance was due to partial obstruction of larger pulmonary vessels, growth in the size of obstructing bubbles could produce a more prominent increase in measured PVR.

In the decompression experiments, He-O₂ breathing was neither beneficial nor deleterious. However, this study did not measure the effect of He-O₂ breathing during recompression. It is possible that He-O₂ breathing might be beneficial when combined with recompression. The lower density of helium as compared to air could conceivably lower the work of breathing and improve gas mixing in the lung. It has been suggested that such an effect might be helpful in cases of DCS with labored breathing (11). We have previously shown that pulmonary (airways) resistance increases to a variable extent in this model of DCS (6). The increases are relatively small, however, and pulmonary vascular obstruction is probably a much more important cause of the observed derangements of gas exchange than is increased airways resistance. Anticipated benefits of He-O₂ breathing at depth due to lowered gas density would need to be weighed against the potential for worsening pulmonary vascular obstruction.

In most cases the major reason for use of He-O₂ breathing during recompression treatments after air dives is likely to be operational convenience. A decision to use He-O₂ in treatment of air DCS must weigh the risk of worsening vascular obstruction, and therefore outcome of treatment. In this study a definite increase in vascular obstruction occurred when ventilation was changed from air to He-O₂. Although lung function did not deteriorate, a similar increase in vascular obstruction might have very deleterious effects if it occurred in other circulations, such as that of the spinal cord. Although it is difficult to predict the effect of gas switching on bubble growth

in other circulations, one animal study has shown that the efficacy of recompression to 100 fsw for the treatment of spinal cord DCS is less with He-O₂ breathing than air breathing (J. J. W. Sykes, personal communication). In view of these experimental observations, it seems reasonable not to use He-O₂ as a breathing gas for treatment of DCS resulting from air dives until its efficacy and safety have been more completely evaluated by experimental and clinical studies.

The authors thank Mr. Todd Dumas and Ms. Patricia Tancos for technical assistance in conducting the experiments, and Mr. Fleetwood Henry for help with the preparation of histologic specimens. We are indebted to Dr. Paul Weathersby, Mr. Randy Hays, and Mrs. Shalini Survanshi for statistical advice, and to Dr. Louis Homer for the computer software used to correct blood gas data. Finally, we thank Ms. Diana Temple for her excellent editorial assistance in the preparation of this manuscript.

This work was supported by the Naval Medical Research and Development Command Research Task No. M0099.PN001.1170. The opinions and assertions contained herein are the private ones of the authors and are not to be construed as official or as reflecting the views of the Navy Department or the naval service at large.—Manuscript received March 1986; revision received August 1986.

Catron PW, Thomas LB, Flynn ET Jr, McDermott JJ, Holt MA. Effets de la respiration de He-O₂ durant la maladie de décompression expérimentale après des plongées à l'air. Undersea Biomed Res 1987; 14(2): 101-111.—Les effets de la ventilation avec He-O2 pendant la maladie de décompression (DCS) et l'embolie gazeuse veineuse, ont été étudiés. Quinze chiens anesthésiés furent ventilés mécaniquement et soumis à des plongées répétées à l'air jusqu'à ce que la pression artérielle pulmonaire augmente d'au moins le double en l'espace de 10 min après la plongée. A 30 min après la plongée, la ventilation fut soit continuée avec de l'air (témoin, n = 7), ou remplacée avec He-O₂ (n = 8) pour une période additionnelle de 90 min. Tous les animaux éprouvèrent une hypertension pulmonaire, hypotension systémique, hémoconcentration, hypoxémie, hypercapnie, et de l'oedème pulmonaire. La respiration d'air ou de He-O₂ après la plongée ne modifia pas ces réponses, mais la respiration de He-O₂ produisit une augmentation de 11% dans la résistance vasculaire pulmonaire (RVP). Chez 3 autres chiens anesthésiés mais qui ne furent pas exposés aux plongées, la ventilation fut changée à l'He-O₂ à des temps variés durant une infusion intraveineuse d'air. La respiration de He-O₂ causa une augmentation de la RVP de 22%. Il est conclu que la respiration de He-O2 durant la DCS résultant d'une plongée à l'air peut aggraver l'obstruction vasculaire pulmonaire.

> plongée maladie de décompression embolie gazeuse veineuse

hélium-oxygène suffocation oedème pulmonaire

animaux

REFERENCES

- 1. Van Der Aue OE, White WA, Jr, Hayter R. Physiologic factors underlying the prevention and treatment of decompression sickness. Research Project X-443, Rep No. 1, April 1945.
- Behnke AR, Jr. Effects of high pressures: Prevention and treatment of compressed air illness. Med Clin North Am 1942; 26:1213-1237.
- 3. Shilling CW. Compressed-air illness. U.S. Naval Med Bull 1938; 36:235-239.
- 4. James PB. The treatment of decompression sickness in air diving. Undersea Biomed Res 1984; 11(Suppl):21.
- 5. U.S. Navy Diving Manual. Vol. 1, Air diving, 1985 Revision.
- 6. Catron PW, Flynn ET, Jr, Yaffe L, et al. Morphological and physiological responses of the lungs of dogs to acute decompression. J Appl Physiol 1984; 57(2):467-474.
- 7. Bove AA, Hallenbeck JM, Elliott DH. Circulatory responses to venous air embolism and decompression sickness in dogs. Undersea Biomed Res 1974; 1:207-220.
- Newman TS, Spragg RG, Wagner PD, Moser K. Cardiopulmonary consequences of decompression stress. Respir Physiol 1980; 41:143-153.

- 9. Sergysels R, Jasper N, Delaunois L, Chang HK, Martin RR. Effect of ventilation with different gas mixtures on experimental lung air embolism. Respir Physiol 1978; 34:329-343.
- Bailey RC, Homer LD. Iterative parameter estimation. Naval Medical Research Institute Technical Rep No. 76-19. Bethesda, MD, 1976.
- 11. U.S. Navy Diving Manual. Vol. 1, Air diving, 1973 Revision.
- James PB. Problem areas in the therapy of neurological decompression sickness. In: Proceedings of a symposium on decompression sickness. North Sea Medical Center, Cambridge, Norwich Union, 1981.
- 13. Van Liew HD, Hlastala MP. Influence of bubble size and blood perfusion on absorption of gas bubbles in tissues. Respir Physiol 1969; 7:111-121.
- Hayduk W, Laudie H. Prediction of diffusion coefficients for non-electrolytes in dilute aqueous solutions. AIChE J 1974; 20(3):611-615.
- Weathersby PK, Homer LD. Solubility of inert gases in biological fluids and tissues: a review. Undersea Biomed Res 1980; 7(4):277-296.
- Munson ES. Effect of nitrous oxide on pulmonary circulation during venous air embolism. Anesth Analgesia 1971; 50(5):785-793.
- Munson ES, Merrick HC. Effect of nitrous oxide on venous air embolism. Anesthesiology 1966; 27(6):783-787.
- 18. Marx GF, Steen SN, Foster ES, Jadwat CM, Kepes ER. Air embolism during neurosurgery. NY State J Med 1968; November:2801-2802.
- Tisovec L, Hamilton WK. Newer considerations in air embolism during operation. JAMA 1967; 201(6):116–117.
- 20. Munson ES, Paul WL, Perry JC, de Padua CB, Rhoton AL. Early detection of venous air embolism using a Swan-Ganz catheter. Anesthesiology 1975; 42(2):223-226.
- 21. Van Liew HD. Dissolved gas washout and bubble absorption in routine decompression. In: Lambertsen, CJ, ed. Undersea physiology IV. Proceedings of the fourth symposium on underwater physiology. New York: Academic Press; 1971:145-150.
- Harvey EN. Physical factors in bubble formation. In: Fulton, JR, ed. Decompression sickness: Caisson sickness, diver's and flier's bends and related syndromes. Philadelphia. PA: Saunders; 1951:90-144.
- Strauss RH, Kunkle TD. Isobaric bubble growth: A consequence of altering atmospheric gas. Science 1974; 186:443-444.
- Powell MR, Spencer MP, Von Ramm O. Ultrasonic surveillance of decompression. In: Bennett PB, Elliott DH, eds. The physiology and medicine of diving. London: Baillière Tindall; 1982;404-434.
- 25. Albertine KH, Wiener-Kronish JP, Koike K, Staube NC. Quantification of damage by air emboli to lung microvessels in anesthetized sheep. J Appl Physiol 1984; 57(5):1360-1368.
- Christman CL, Catron PW, Flynn ET, Weathersby PK. In vivo microbubble detection in decompression sickness using a second harmonic resonant bubble detector. Undersea Biomed Res 1986; 13:1-18.

JECONT CLASSICATION OF THE PARTY	REPORT DOCUM	MENTATION	PAGE				
14. REPORT SECURITY CLASSIFICATION	16. RESTRICTIVE MARKINGS						
Unclassified							
28. SECURITY CLASSIFICATION AUTHORITY		3. DISTRIBUTION / AVAILABILITY OF REPORT					
2b. DECLASSIFICATION / DOWNGRADING SCHEDULE		Approved for public release;					
4. PERFORMING ORGANIZATION REPORT NUM NMRI 87-15	BER(S)	S. MONITORING	ORGANIZATION	REPORT NUMBER	s)		
6a NAME OF PERFORMING ORGANIZATION Naval Medical Research	6b. OFFICE SYMBOL (If applicable)	7a. NAME OF MONITORING ORGANIZATION Naval Medical Command					
6c. ADDRESS (City, State, and ZIP Code)		7b. ADDRESS (City, State, and ZIP Code)					
Bethesda, Maryland 20814-5055	Department of the Navy						
Deciredad, 122/22112 2007 3003		Washington, D.C. 20372-5120					
8a. NAME OF FUNDING/SPONSORING ORGANIZATION Naval Medical (If applicable) Research and Development Command		9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER					
BC. ADDRESS (City, State, and ZIP Code)		10. SOURCE OF FUNDING NUMBERS					
Bethesda, Maryland 20814-5055		PROGRAM ELEMENT NO. 63713N	PROJECT NO. MO09.01A	TASK NO. 01A-0003	WORK UNIT ACCESSION NO. DN777317		
		03/130	1007.018	0111 0000	וואען אוען		
REPORT NO. 8 FROM_ 16. SUPPLEMENTARY NOTATION	TO	14. DATE OF REPO	·	n, Day) 15. PAGE 11 ch 1987 pp			
17. COSATI CODES	I IR CURIECT TERMS	18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)					
FIELD GROUP SUB-GROUP	FIELD GROUP SUB-GROUP diving: dec			compression sickness; Venous air embolism			
	animals: he	animals; helium-oxygen; chokes; pulmonary edema					
					·		
19. ABSTRACT (Continue on reverse if necessa	ry and identify by block	number)					
			•				
					•		
,	•	•			•		
	-						
					•		
•					•		
,							
•							
~ .							
20. DISTRIBUTION/AVAILABILITY OF ABSTRA	_		ECURITY CLASSII	FICATION	· · · · · · · · · · · · · · · · · · ·		
☐ UNCLASSIFIED/UNLIMITED ☐ SAME A	IS RPT. DTIC USERS			and the order	CYMAROL		
224. NAME OF RESPONSIBLE INDIVIDUAL Phyllis Blum, Information S	ervices Division	202-295-218	(include Area Co 38	ISD/ADM	ÍN/NMRI		